



# Using GOALIE to Analyze Time-course Expression Data and Reconstruct Kripke Structures

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# Outline

- Interactions between experiments, data and interpretation
- Models of Biological Processes and Systems
  - Description (via controlled vocabularies and ontologies)
  - Reconstruction (via time-course analysis and statistical procedures)
  - Model Repositories
- Computational “Searches” for “models” (parameters, new interactions, etc)
  - Problems
    - Low sampling rate
    - Upsampling, optimization schemes
    - Models limitations



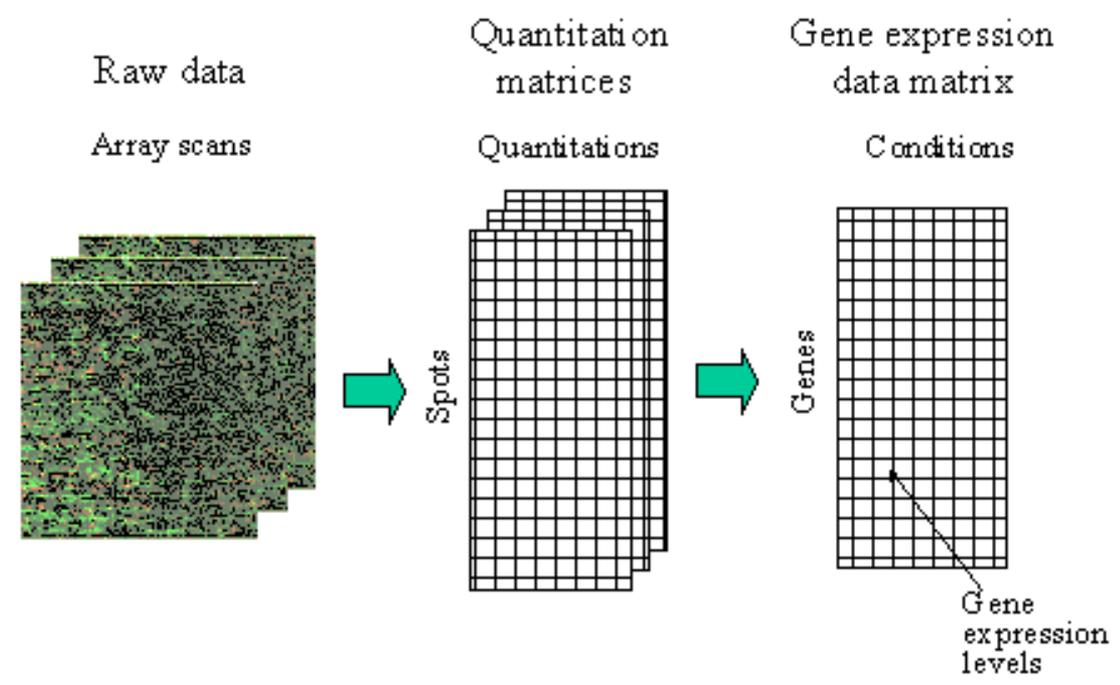
# Analyzing Time-course Microarray Experiments

- Microarray Experiments and Data
- “Enrichment” studies via Controlled Vocabularies and Ontologies (Gene Ontology and others)
- Model “reconstruction”
  - Similarity studies
  - Segmentation algorithms
  - Kernel methods
  - Results
- Future work
- Joint work with Bud Mishra, Courant NYU, Naren Ramakrishnan, Virginia Tech, Daniele Merico, University of Toronto, many others at NYU and UNIMIB



# Microarray Experiments

- From laser scans readings, a numerical value corresponding to the relative expression of a "gene" is produced.
- When each raw data array scan corresponds to a given time-point under a specific condition, the final gene expression data matrix represents the temporal evolution of the gene expression.





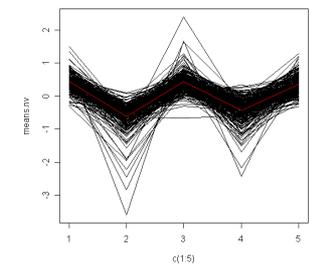
# Standard data-mining approaches to microarray data

- The results of microarray experiments have been studied by means of statistical techniques
- Aim:
  - To group together genes/probes that “behave similarly” under different experimental conditions (usually achieved by *clustering*)
- Successful endeavor
  - Several tools and libraries are provided to perform this kind of studies
  - Several publications produced with results in this field
  - Many of the studies reported still contain a considerable amount of “hand curation”

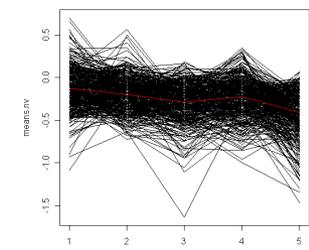


# Standard data-mining approaches to microarray data

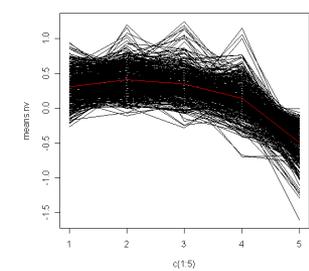
- The expression matrix is usually analyzed according to standard techniques:
  - Clustering  
enables to group together genes with a similar expression profile
  - Gene Ontology (GO) terms “Enrichment”  
enables to find statistically over-represented terms in given set of genes - i.e., clusters - thus providing some “functional” characterization
    - usually computed using some *statistical significance test*; e.g., Fisher’s exact test, Hypergeometric Test, Binomial Test,  $\chi^2$  Test, plus various corrections



- Ribosome  
- Translation



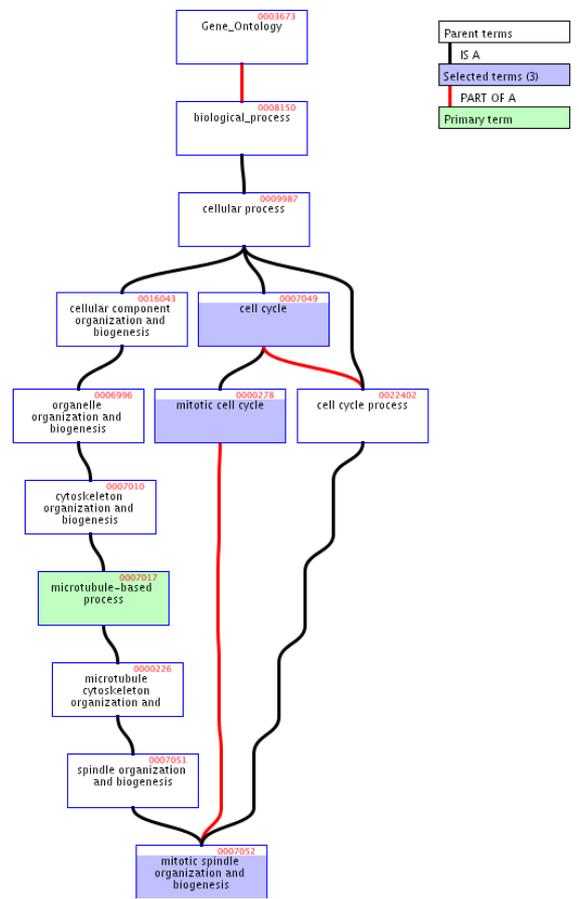
- Spindle  
- Cell wall  
- Budding



- Glucose Transport



# Gene Ontology (GO)



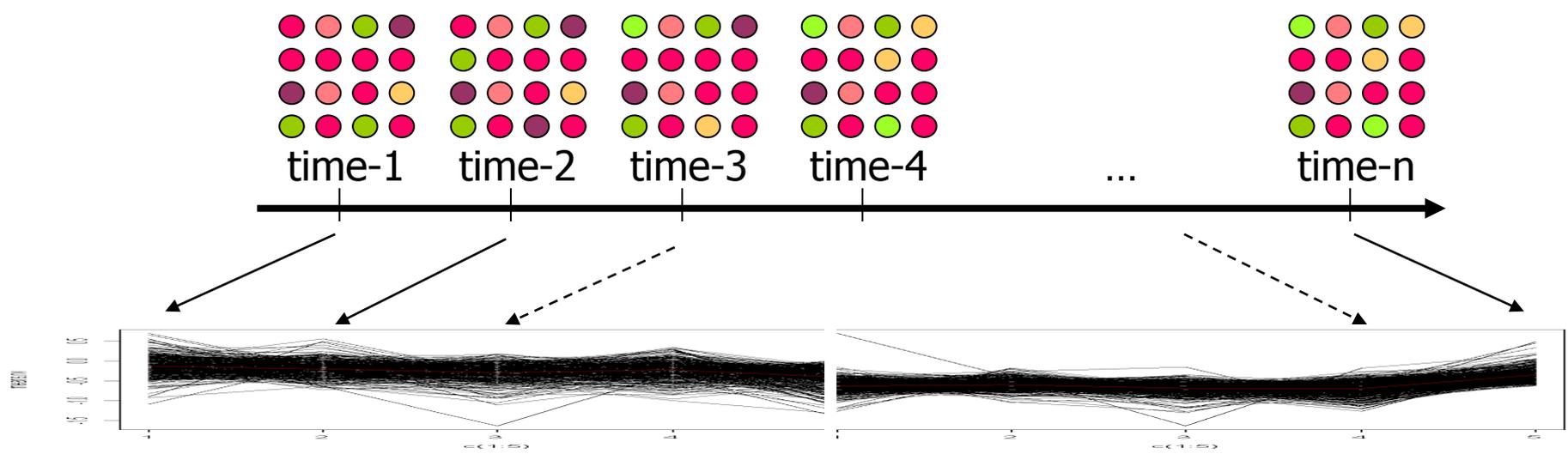
- GO is a controlled vocabulary for the functional annotation of genes
- GO is composed by three independent classifications, each of them having a hierarchical DAG structure
  - **MF**: Molecular Function (biochemical activity and molecule type)
  - **BP**: Biological Process
  - **CC**: Cellular Component

[www.geneontology.org](http://www.geneontology.org)



# Time-course microarray data

- Clustering is performed with all time-points together spanning the whole time-course



- This amounts to assume that if genes are co-regulated across **some** time-points, they will also be co-regulated throughout the **whole** time-course
- However, co-regulation may be interrupted at a certain point
  - Different short-time and long-time response, e.g., *DNA damage*
  - Multiple-stages transcriptional program, e.g., *development*



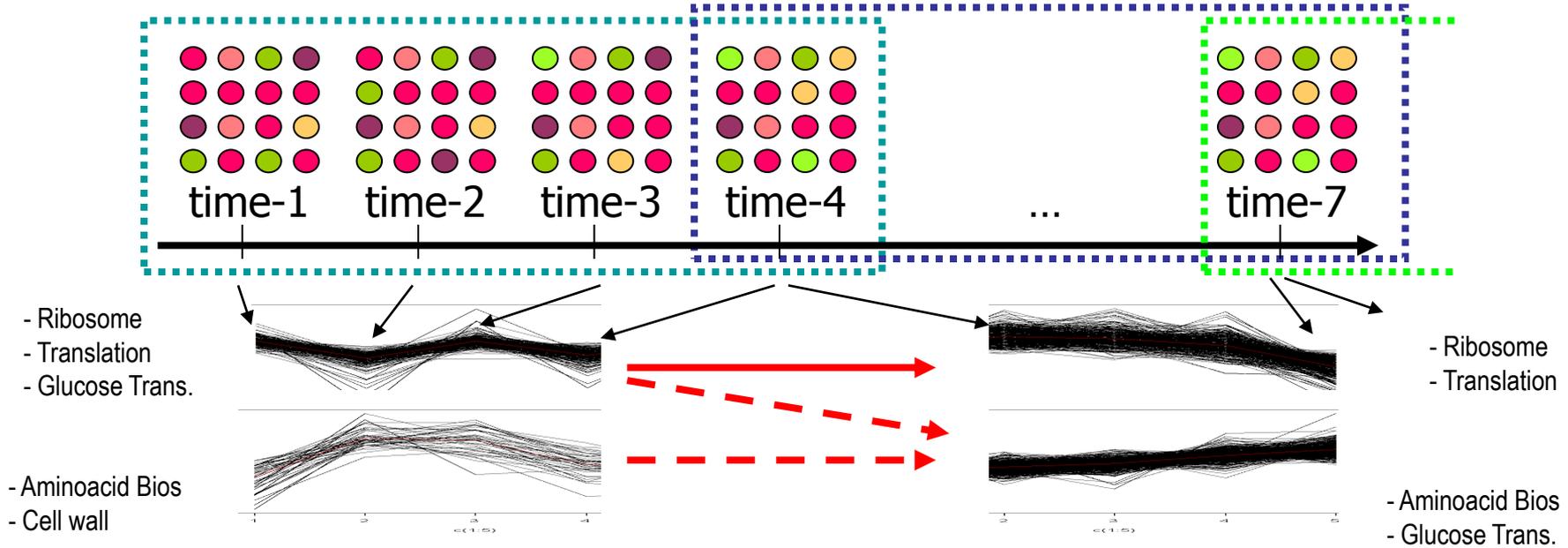
# GOALIE: a twist on “enrichment” studies

- **GOALIE** introduces a twist on enrichment studies by taking into account possible temporal variations of biological processes in time-course measurements
- The key observation is that an “enrichment” of a set of genes/probes may vary depending on the length of the (time) vector of measurements
- **GOALIE** assumes that the a time-course experiment has been broken down into windows and that each window has been clustered separately
- Afterward the enrichment of each cluster in a window is compared with the enrichment of clusters in neighboring windows and all the possible relations are built in a DAG
  - **GOALIE** provides several interfaces to explore, summarize and compare the DAGs pertaining to different experiments



# Piece-wise approach to time-course microarray data

- We split the time-course into discrete windows,
- Then compute clusters for each window separately,
- Finally reconnect clusters from adjacent windows exploiting similarity of Gene Ontology cluster enrichments





# Computational Modules

- In order to enhance the GOALIE software we concentrated on the components computational modules
- Computational modules are required for:
  1. Clustering (*Clique* [Shamir et al.], K-means, SVM, SOMs etc.; tool *Genesis* from TU-Graz and many other ones)
  2. Segmentation (PNAS 2010 [Ramakrishnan et al.]
  3. Gene Ontology (GO) enrichment (Fisher's exact test etc.)
  4. Computing similarity among clusters from adjacent time-windows, based on GO enrichment (*ex-novo* – Kernel function)
  5. Select only relevant connections among clusters (*ex-novo*)
- In the rest of this presentation, the focus will be on the Kernel approach developed for module #4; #5 has been published in (CaOR 2010 [Antoniotti et al.]



# Computing “Similarity” Using Graph Kernels

- The results of the first three steps of the algorithm consist in the “enrichment” of each cluster by a set of representative labels (GO terms)
- Next we want to see how similar two clusters are based on this labeling
- **Note**
  - This check may be useful to a biologist trying to track biological processes over time; e.g., trying to see which genes are involved in a certain process as time evolves
  - From a more abstract point of view this is a procedure that measures how two objects are similar
    - The similarity between the two objects is done in a **re-described** space (possibly with lower dimensionality)
    - In our case there is some more structure we want to exploit



# Computing “Similarity” Using Graph Kernels

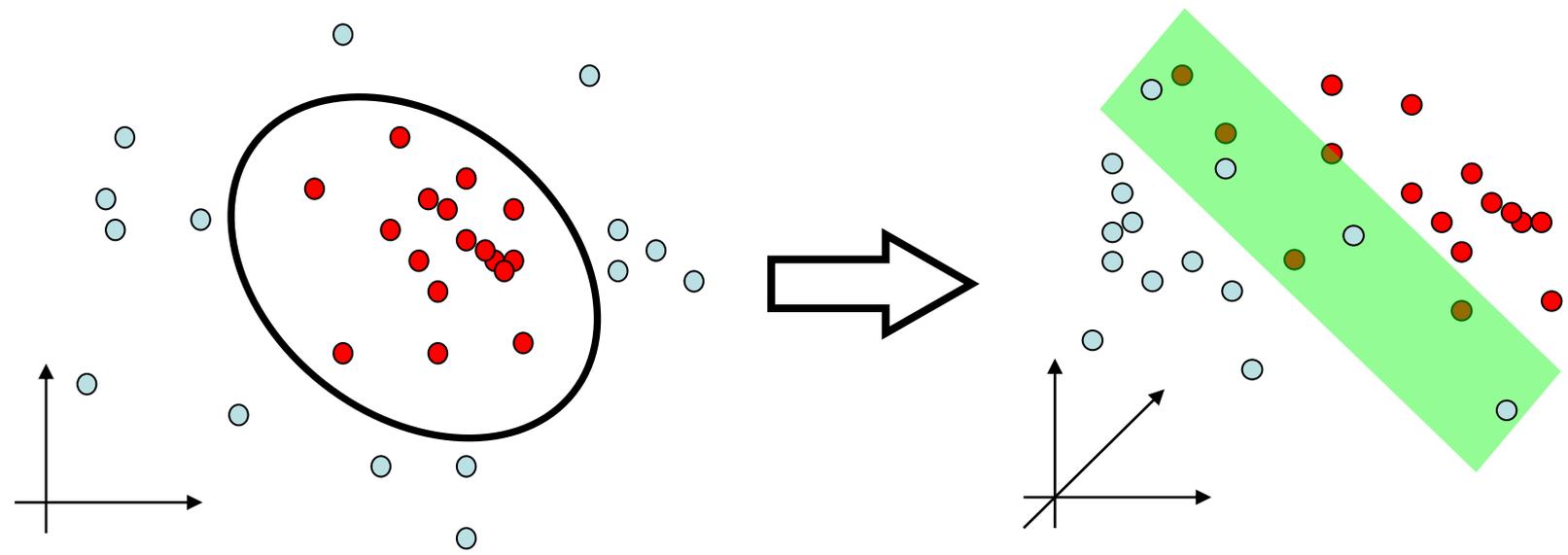
- Peculiarities of our method
    - Our objects are clusters ordered in a time-course
    - The labeling by GO terms does have a structure imposed by their hierarchical arrangement in a DAG
  
  - Previous work
    - Similarity between objects of this kind is computed using various measures
    - In the specific case of labeling of gene sets, flat lists of symbols were used
      - Similarity computed Jaccard index
- $$J(X, Y) = 1 - \frac{|X \cap Y|}{|X \cup Y|}$$
- Graph kernels can instead be used to take into account the DAG nature of the GO labels
    - Question: what is the performance of our Graph Kernel method w.r.t. a simple Jaccard index calculation?



# Kernel Methods

When the existence of a non-linear pattern prevents from using a linear classification algorithm, the problem can be solved introducing a mapping function  $\Phi$  which projects the problem in a higher dimension space, where the pattern is linear

$$\Phi : R^N \rightarrow R^M \quad (M > N)$$





# Kernel methods

- How to perform the mapping?
  - We don't really have to know the mapping  $\Phi$  if we introduce a **Kernel function  $k$**

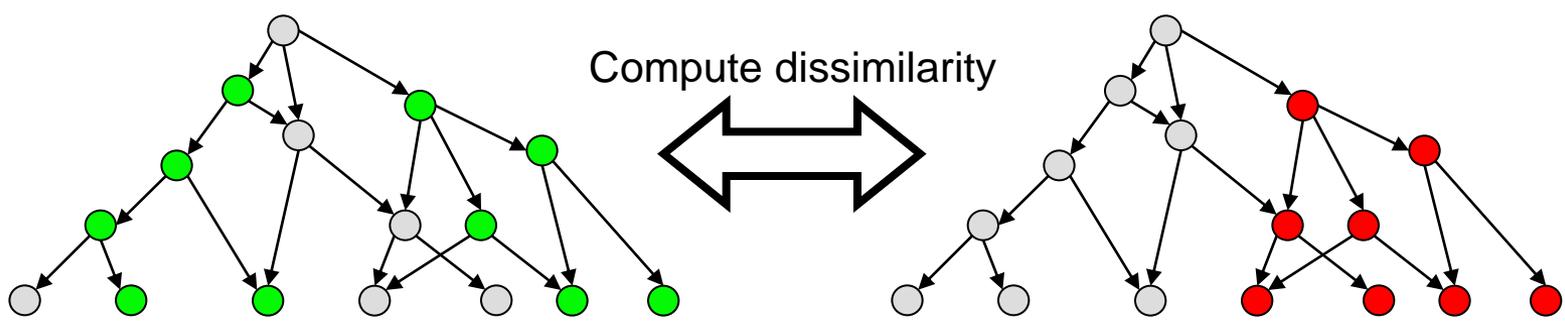
$$k(x, y) = \langle \phi(x), \phi(y) \rangle_F$$

- The internal product between the remapped points is compute by  $k$  thus avoiding the explicit computation of  $\Phi$  (the so called **Kernel Trick**)
- In order to be a proper Kernel, a function must be positive semi-definite and symmetric (Mercer's Theorem)
- A Kernel function can also be used to induce a dissimilarity function (that's exactly what we do)



# A Kernel Function for Gene Ontology Graph Comparison

- Input: GO enrichment graph; i.e., sub-graphs of the overall GO taxonomy for each cluster
  - Each vertex is identified by a **label** - the GO term name - which is then used for walk matching
  - Each vertex has also an associated **p-value** label, from Fisher's exact test, which is then used to compute a dissimilarity score between the walks
    - We work on GO sub-graphs (forests), obtained by filtering in only the terms with  $p\text{-value} < \text{significance threshold}$

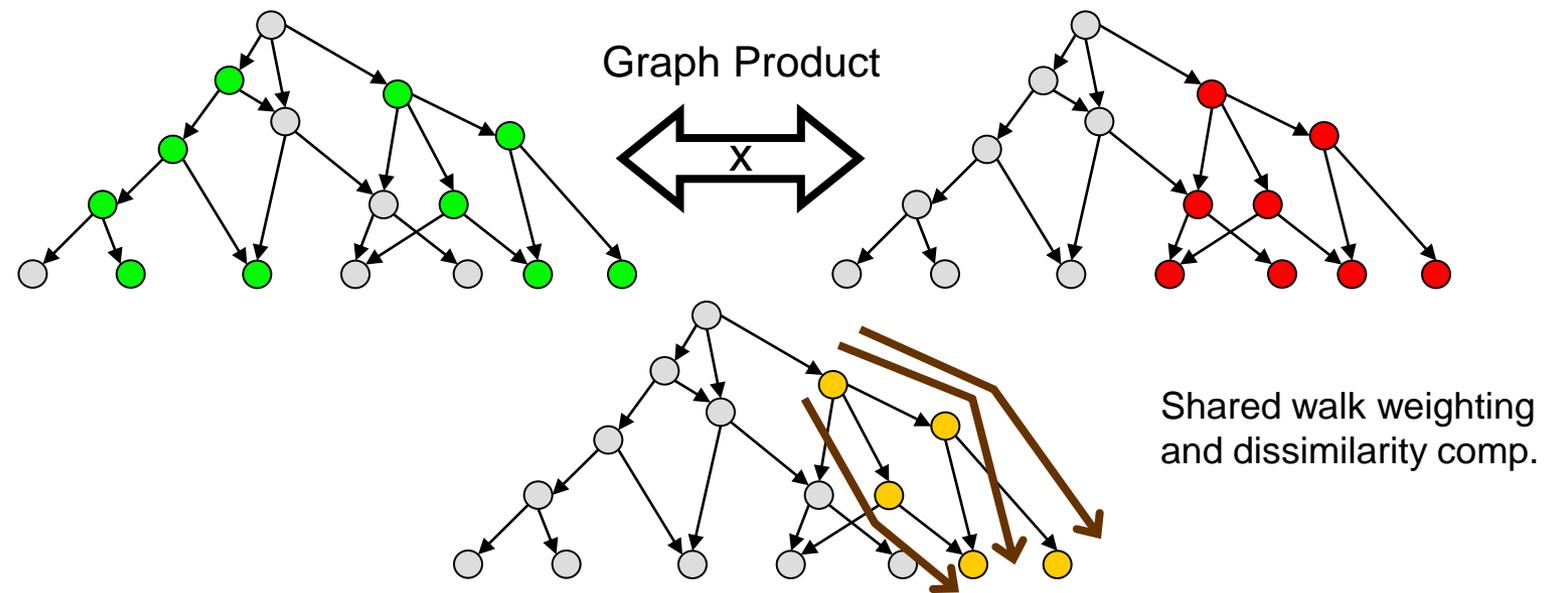


Colored dots represent GO terms with  $p\text{-value} < \text{significance threshold}$



# A Kernel Function for Gene Ontology Graph Comparison

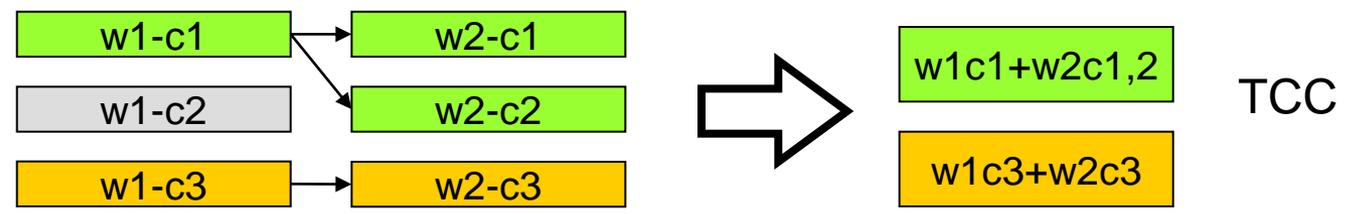
- The computation (informally) proceeds in the following way
  - We compute the (direct) **graph product** between the two GO sub-graphs
  - We identify **common walks** in the product GO sub-graph
  - We compute a weighted **dissimilarity** score for each walk
  - We sum all the walk dissimilarities to get the **total dissimilarity**





# A Kernel function for Gene Ontology graph comparison

- What are the advantages of our approach?
  - We explicitly take into account the hierarchical structure of GO cluster enrichments (Zoppis et al. 07 ISBRA)
- Next we concentrated on evaluating our approach
  - For a benchmark for our Kernel function we set up a comparison with a Jaccard Coefficient-based dissimilarity, working on GO enrichments as flat lists of terms
    - Once the dissimilarities are computed with both methods, we select only significant similarity patterns among clusters from adjacent windows (\*)
  - We also consider a model manually curated by an expert
  - To quantitatively assess performance, we adopt the Loganantharaj et al (BMC Bioinformatics, 2006) **Total Cluster Cohesiveness** (TCC) score, which enables to assess the homogeneity of a cluster in terms of its GO terms; we compute TCC for groups of connected clusters (Merico et al. 07 KES-WIRN)



# GOALIE Interface

The screenshot displays the GOALIE interface with the following components:

- Clusters Connections:** A hierarchical tree diagram showing clusters like 'alpha-42-70-cluster-10' and 'alpha-63-91-cluster-14' connected to various gene clusters (e.g., 13, 20, 2, 8, 5, 1, 1, 6, 4, 1, 1, 4).
- Clusters Information:** A table with three columns: 'Cease to be active', 'Remain active', and 'Become active'. It lists GO terms such as 'ribosomal large subunit assembly', 'mitotic sister chromatid segregation', 'RNA-nucleus', 'nucleocytoplasmic transition', 'purine nucleoside triphosphate biosynthesis', etc.
- Current Selections:** A panel showing 'Selected Items' (YHR166C, YBL023C) and 'Selected Terms' (mitotic spindle elongation, chromosome segregation, myo-inositol metabolism).

Clusters connection tree  
Each level a "window"

Micro-array accessions

GO categories

Clusters information

Connection information

Cluster Information

# GOALIE Interface

The screenshot displays the GOALIE interface with the following components:

- Source cluster:** A table listing gene accessions and their associated GO categories.
- GO Categories:** A list of GO terms and their descriptions for the source cluster.
- Edge 2 ==> 15:** A comparison table between source and destination clusters.
- Edge cover:** A list of GO terms shared between source and destination clusters.
- Becomes true:** A list of GO terms present in the destination cluster but not in the source cluster.
- Cease to be true:** A list of GO terms present in the source cluster but not in the destination cluster.
- Dest:** A list of GO terms for the destination cluster.

Annotations in blue boxes explain the categories:

- GO categories describing genes in "source" cluster:** Points to the 'GO Categories' list.
- GO categories shared with "destination" cluster:** Points to the 'Edge cover' list.
- GO categories describing "destination" cluster but not "source":** Points to the 'Becomes true' list.
- GO categories describing "source" cluster but not "destination":** Points to the 'Cease to be true' list.



# GOALIE Interface

**Spellman Yeast Cell Cycle Data - Alpha**

File Views Help

**Clusters Connections**

Gantt Chart View | Graph View

Show all terms | Show selected terms

establishment and/or maintenance of chromatin architecture  
G1 phase of mitotic cell cycle  
mitotic spindle elongation  
chromosome segregation  
myo-inositol metabolism

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**Clusters Information** | **Current Selections**

Clusters information | Connection information

Edge "alpha-42-70-cluster-10" ==> "alpha-63-91-cluster-14"

Cease to be active | Remain active | Become active

- Cease to be active:**
  - R ribosomal large subunit assembl
  - R mitotic sister chromatid segreg
  - R mitotic anaphase
  - R G1-specific transcription in mit
  - I regulation of transcription f
  - P G1 phase of mitotic cell cy
  - R poly(A) tail shortening
  - R meiotic DNA recombinase ass
  - R double-strand break repair via
  - R recombinational repair
  - R DNA recombinase assembly
  - R myo-inositol metabolism
  - R transcription initiation
- Remain active:**
  - R transcription m
  - R RNA-nucleus
  - R nucleocytopla
  - R metabolism
  - R regulation of c
  - R nucleotide me
  - R purine ribonuc
  - R nucleotide bio
  - R ribonucleotide
  - R amine metabo
  - R cytoplasmic tr
  - R histone acetyl
  - R actin filament
  - R negative regu
- Become active:**
  - R purine nucleoside triphosphate biosynthesis
  - R purine ribonucleotide biosynthesis
  - R ribonucleoside triphosphate metabolism
  - R ribonucleoside triphosphate biosynthesis
  - R purine ribonucleoside triphosphate metabolism
  - R purine ribonucleoside triphosphate biosynthesi
  - R ribonucleotide biosynthesis
  - R amine catabolism
  - R response to external stimulus
  - R chromatin modification
  - I establishment and/or maintenance of chro
    - I DNA packaging
    - I chromosome organization and biogene
  - R ethanol fermentation

**Current Selections**

Selected Items

YHR166C  
YBL023C

Selected Terms

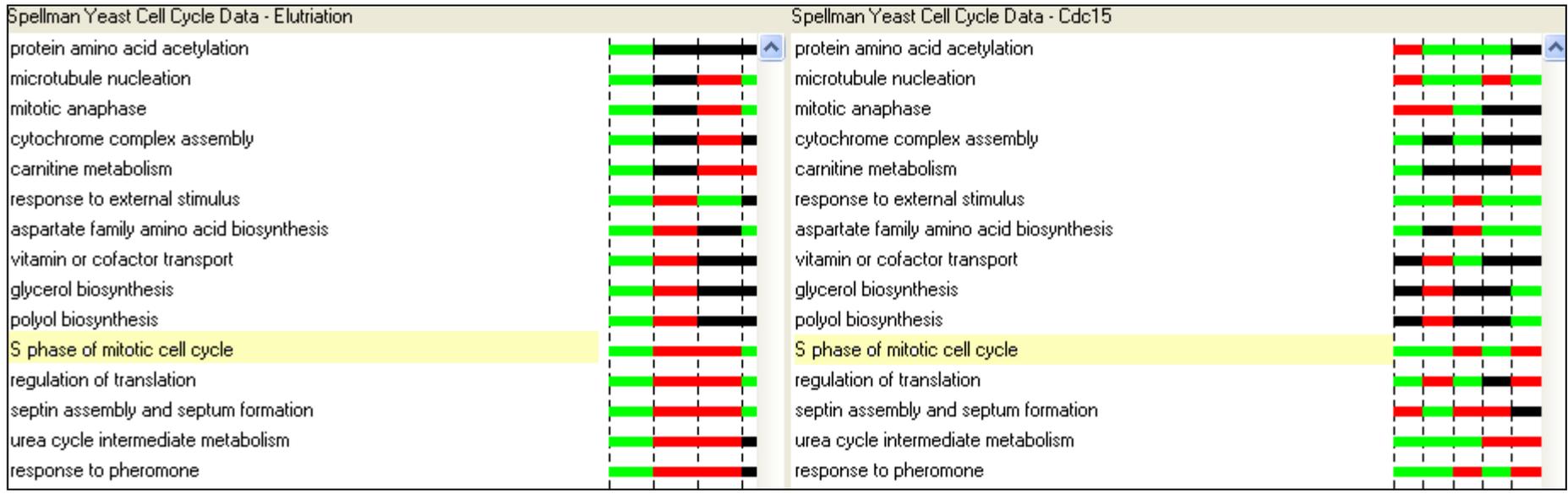
establishment and/or maintenance of  
G1 phase of mitotic cell cycle  
mitotic spindle elongation  
chromosome segregation  
myo-inositol metabolism

Clear selections | Clear all

Status: Selected Terms Structure: Gene Ontology, Computation Method: (FTBH with parameter 0.05)



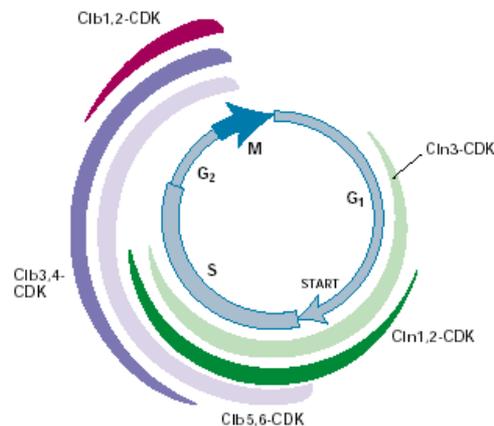
# GOALIE Interface



GOALIE summary comparison view of two cell cycle experiments



# Yeast Cell Cycle benchmark



- Cell Cycle is a multi-stage phenomenon (phases), therefore co-regulation patterns may change across time
  - In [Ramakrishnan et al. 2010] we consider different datasets regarding YCC and Yeast Metabolic Cycle
  - In particular, we consider two windows: G<sub>1</sub>>S and G<sub>2</sub>>M>G<sub>1</sub>
- We use Spellman microarray yeast cell cycle data (1998; a well known benchmark for testing novel analysis tools and methods)
  - CDC15-mutant synchronization
  - ALPHA factor synchronization



# Comparison results using KL segmentation

Yeast  
“Metabolic”  
Cycle  
Segmentation  
Comparison:  
8 segments  
inferred

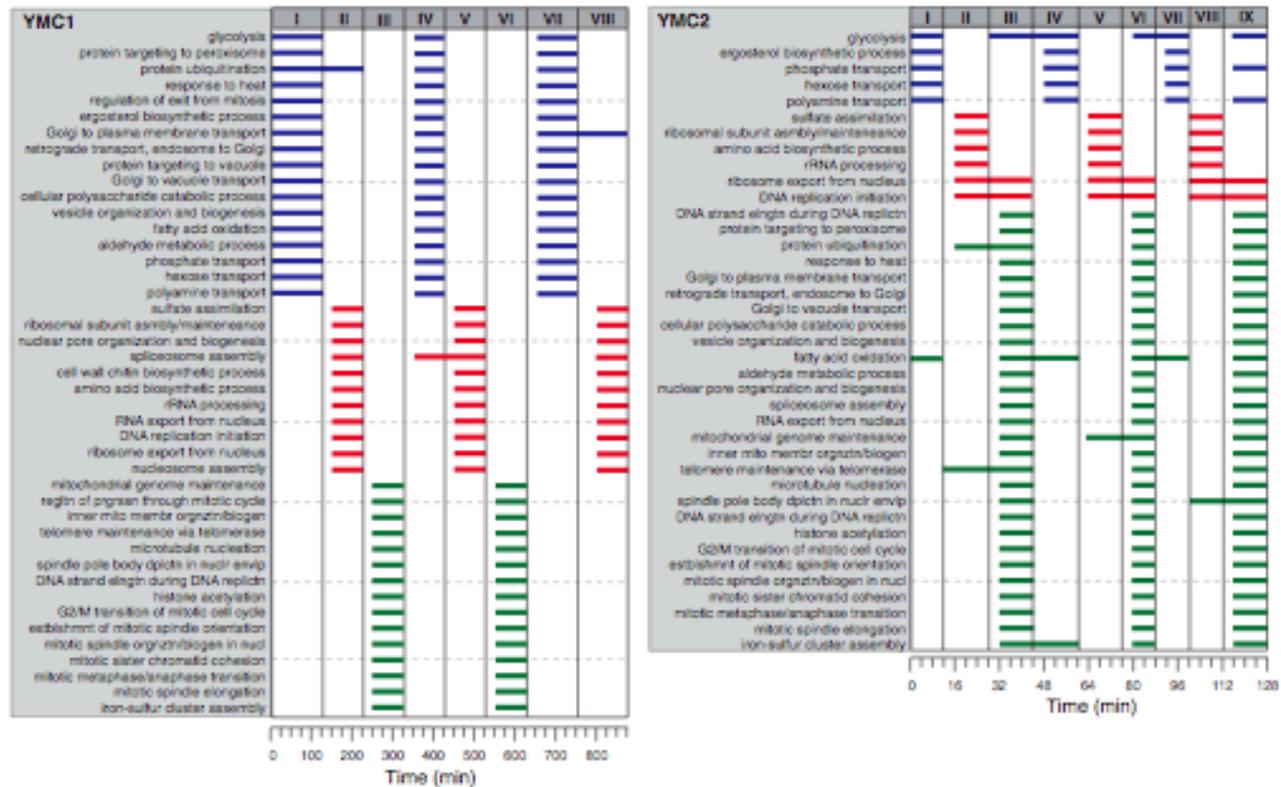
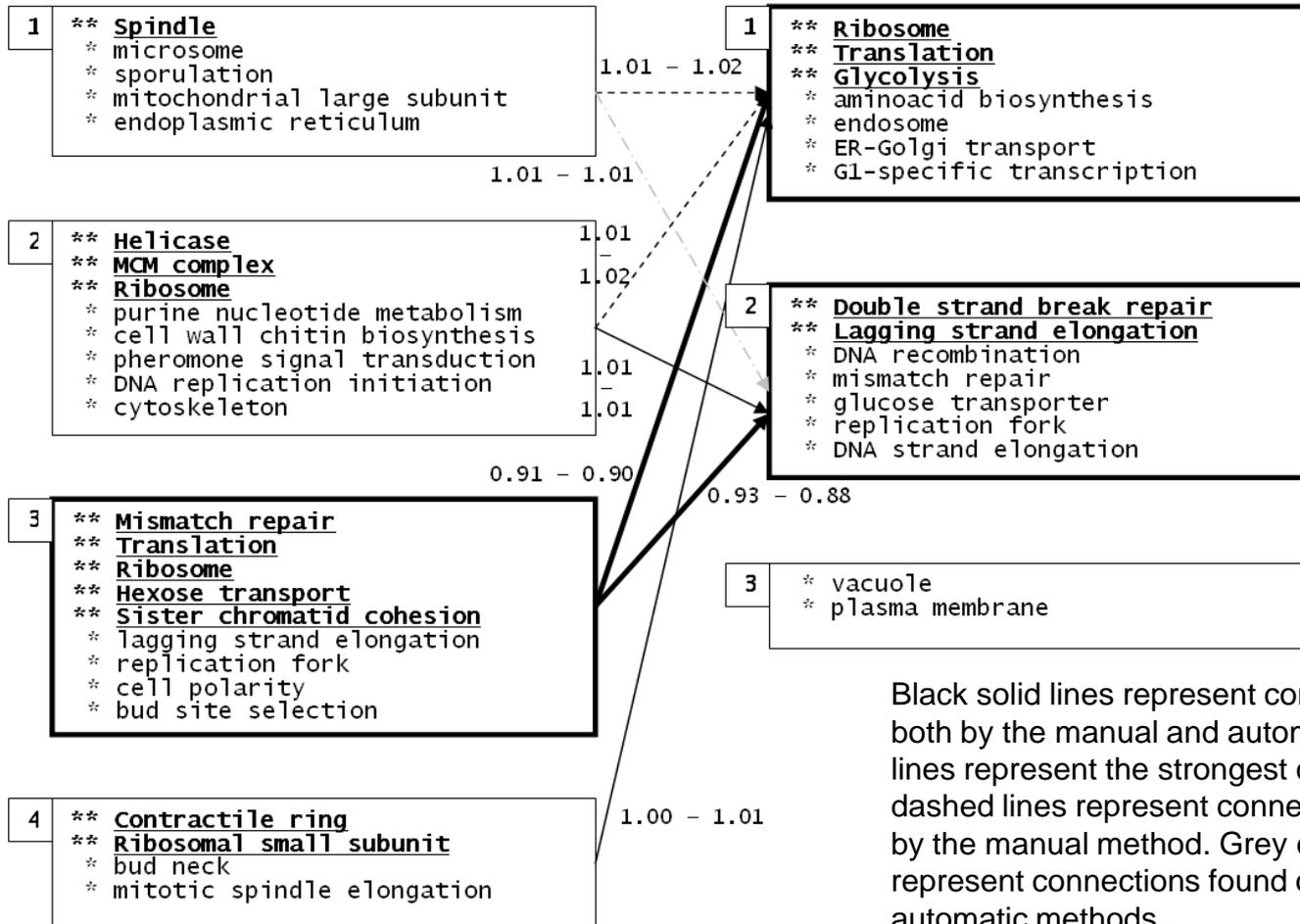


Fig. 4. Segmentation resulting from the GOALIE analysis of transcriptional profiling datasets evaluating the rhythmical growth of *S. cerevisiae* (YMC1: diploid CEN.PK122, nutrient-limited conditions; YMC2: diploid IFO0233, not nutrient limited). The time line of each experiment is shown with each hash mark indicating a sampling point. GOALIE accurately determined the G1, S, and G2/M phases of the cell cycle, respectively. Note that the genes associated with each segment were culture and strain-dependent.



# Results

Inferred cluster connections



Black solid lines represent connections found both by the manual and automatic methods; Bold lines represent the strongest connections. Black dashed lines represent connections found only by the manual method. Grey dash-dotted lines represent connections found only by the automatic methods..



# Results

## Results overview

- Main results were generated for Alpha subset (2 windows), displaying a substantial convergence between the three methods
  - Numerical results are comparable with Jaccard method
  - Kernel method is more “correct” from the information point of view
  - Kernel method is more computationally intensive
- Preliminary results were also generated for CDC15 subset, displaying a better performance of Kernel over Jaccard

## Results (Alpha subset)

<b>Distance</b>	<b>TCC</b>	<b>threshold</b>
Jaccard	94.28	0.05
Jaccard	92.95	0.01
Jaccard	92.95	0.005
Kernel	92.95	0.01
Kernel	94.63	0.05
Manual	92.27	N/A



# Problems

- **Low sampling rate:** biological experiments usually have a way too low sampling rate
  - **Ok** for long term observations at equilibrium
  - **Not ok** for transients and discontinuities detection
    - **Assumption:** transients and discontinuities are interesting
- **Solutions**
  - **Upsampling** after fitting the data to a set of interpolating functions (rational functions or polynomials)
  - Merging of different data sources
    - Several institutions and databanks (e.g., GEO) contain several experiments
    - “Related” experiments can be combined to yield a **Virtual Time-Course Experiment** that organized the extant corpus of knowledge



# Current and future research

- Connection ordering between clusters
  - Method based on optimization of (average) entropy orders connections according to a decrease in the uncertainty of the result graph Kernel similarity between the labeling of two clusters (Antoniotti et al. CaOR 2010)
    - “Complementary” with work on segmentan based on KL divergence published in Ramakrishnan et al. PNAS 2010
- Sample classification (i.e. VTE reconstruction) can be performed if there is an appropriate model of the underlying biological system
  - Ontology research
    - Signs Symptoms Findings Workshop in Milan, 3-4 September 2009



# Current and future research

- **Temporal Series Reconstruction** is a hard problem (deterministically akin to the Traveling Salesman Problem)
  - Bar-Joseph models based on EM optimization procedure
  - Magwene and Kim procedure based on heuristic MST built on top of PQ-trees
  - Lack of data points is a problem
- **Prediction Models**
  - What happens if we “extend” a time course in the future?



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Thank you!